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# REVIEW

# Transcription Factor NF-κβ and Molecules Derived from its Activation in Age-Related Macular Degeneration

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# ABSTRACT

Activation of the nuclear factor kappa  $\beta$  (NF- $\kappa\beta$ ) is related to many inflammatory diseases, including age-related macular degeneration (AMD). The imbalance in the redox state, which happens mainly in senescence, associated with several peculiar characteristics of the macular region, has led to studies of this molecule for AMD therapeutic interventions. Findings report the involvement of NF- $\kappa\beta$  both in the triggering as well as in the worsening condition of AMD. The present article correlates AMD oxidant and inflammatory genesis with the action of the nuclear factor kappa  $\beta$ . Besides its mechanism of action, this study also analyzes the main inflammatory cytokines and adhesion molecules that may be activated by NF- $\kappa\beta$  and are closely related to AMD

KEYWORDS: Macular Degeneration; NF-κβ; Oxidation; Inflammation; Cytokines

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# INTRODUCTION

Age-related macular degeneration is the main cause of irreversible loss of vision in the elderly in developed countries (1,2). Although AMD physiopathogenic mechanisms are not completely explained, some peculiarities of the macular region that induce its degeneration have already been established. The retina is a tissue exposed to oxidative stress due to its high metabolism, large concentrations of polyunsaturated fatty acid content, exposure to visible light (between 400 - 700 nm) and the presence of photosensitive molecules such as rhodopsin and lipofuscin (3). The oxidative and nitrosative stress to which the retina is exposed is induced by the imbalance between the antioxidant defense and the production of reactive oxygen species (ROS) and reactive nitrogen species (RNS) and plays an important role in the triggering and progression of AMD (3-5). Photosensitive reactions, for example, generate ROS and RNS, such as superoxide (O2<sup>-</sup> •), hydrogen peroxide (H2O2), singlet oxygen (102), and peroxynitrite (ONOO-), which induce damage to retinal pigment epithelial (RPE) cells (6,7). The hypofunctioning RPE cells inhibit the appropriate degradation of the products resulting from the phagocytosis of the photoreceptor outer segment cells,

causing the pathological accumulation of lipids in the Bruch's membrane (8,9), producing druses and other extracellular deposits in the Bruch's membrane. These deposits are considered important risk factors for the development of AMD (8,9). The druses, as well as the choriocapillaris, the photoreceptors and the RPE cells, present inflammatory and immunological markers (10-22). Additionally, microglia, the immune cells responsible for the coordination of responses to inflammatory stimuli of the retina (23-24), as well as the RPE cells and the macrophages, secrete cytokines, enzymes, and growth factors, responsible for the triggering and the progression of AMD (25-28).

Inflammation is an important activator of the nuclear factor kappa  $\beta$  (NF- $\kappa\beta$ ). When activated, this transcription factor induces an increase in inflammatory cells and molecules perpetuating the cycle (29). Additionally, NF- $\kappa\beta$  is also a redox-sensitive transcription factor, that is, its activation is triggered by the cell oxidative stress (30-35). Several studies correlate NF- $\kappa\beta$  with AMD (36-39). Hence, this review discusses the role of the nuclear factor kappa  $\beta$  (NF- $\kappa\beta$ ) and its activated inflammatory molecules in AMD genesis.

# NF-kβ

Transcription factors are proteins responsible for the coordinated expression of genes through specific binding to gene promoter and enhancer sites (40). NF- $\kappa\beta$ transcription factor was discovered in 1986. It was first identified in T lymphocytes, and later observed in all mammal cells (41-42). It plays an important role in the cell survival and proliferation, as well as in its apoptosis (43-44). Additionally, it helps to regulate the expression of genes associated with the immune and inflammatory responses (45-46). It is important to note that this transcription factor mediates the synthesis of cytokines such as tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), Interleukin-1 $\beta$ (IL-1β), Interleukin-2 (IL-2), Interleukin-6 (IL-6), and Interleukin-8 (IL-8), as well as the expression of the cyclooxygenase 2 (COX-2), inducible nitric oxide synthase (iNOS), and acute phase proteins, such as creactive protein (CRP). Besides the response it provides to the acute inflammation, NF- $\kappa\beta$  is a master regulator of the chronic inflammatory processes (47-48). Such cytokines may cause oxidative stress-induced cell dysfunction or cell death (49). NF- $\kappa\beta$  activation is also related to the increase in the expression of the adhesion E-selectin, intercellular adhesion molecule-1 (VCAM-1), and vascular cell adhesion molecule-1 (ICAM-1), whereas inhibition of NF- $\kappa\beta$  decreases the transmigration and the leukocyte adhesion (50).

NF-κβ family (or Rel family) is composed of five subunits: p65 (RelA), c- Rel, RelB, p50 and p52. It is characterized by including a well-preserved N-terminal domain with around 300 amino acids (RHD – *Rel homology domain*), which subdivides into two domains, the DNA-binding and the dimerization one (43, 51-53). NF-κβ subunits homo- or hetero-dimerize to form activating dimers (p50-p65) or repressors (p50-p50 e p52-p52). They are found in the cytoplasm of most cells in an inactivate state, binding with the inhibitory proteins of the inhibitory kappa B (IκB) family, among which the most important are IκBα, IκBβ, and IκBε. IκBα is associated with the transient activation of NF-κβ, whereas IκBβ is involved in sustaining the activation (54-57).

There are two pathways to activate the NF-κβ: the classical (canonical) and the alternative (non-canonical) pathways. The classical one is more common and is associated with genes, innate immunological inflammation-related response, anti-apoptosis and cell survival (40). Conversely, the alternative pathway is associated with the expressions of genes that contribute to develop and maintain the secondary lymphoid organs (58). When not activated, NF- $\kappa\beta$  factor is found in the cytoplasm, binding to an inhibitory protein, IkB. This complex prevents the translocation of NF-κβ into the nucleus. Hence, IkB phosphorylation and degradation are required for translocation to occur (43-44,54,59). After IkB degradation, NF-KB dimers (E.g. p50-p65) are released and migrate into the nucleus where they will bind with kB target gene enhancers, inducing the transcription of genes that mediate several cellular processes such as immunity, inflammation, proliferation, apoptosis and cellular senescence (60-62).

Several internal and external cell stimuli may contribute to this activation, such as neurotrophins, neurotoxic proteins (such as  $\beta$ -amyloid), cytokines (Interleukin-1 and TNF- $\alpha$ ), glucocorticoid, phorbol esters, atrial natriuretic peptide,

ceramides, virus- and bacteria-derived products, ultraviolet irradiation, ionizing radiation, enzyme reaction products such as iNOS and COX-2 (29,47,63-66). It is important to point out that during chronic inflammation, several immunological cells are continuously activated by inflammatory mediators, and when inflammation is not resolved, the cells recruited by the inflammatory mediators secrete additional mediators, inducing a vicious cycle that activates NF- $\kappa\beta$  in a chronic way (29).

# THE ROLE OF NF-κβ IN AMD PATHOGENESIS

It is possible to infer that, due to the fact that NF- $\kappa\beta$  is activated by the oxidative stress and the inflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$ , as well as by the concentration of UV rays, the macular region meets the appropriate conditions for its activation (63-64,67-68). Besides these factors, the chronic oxidative stress induces the production of advanced glycation end products (AGE) and their receptors (RAGE). It is important to point out that increased RAGE or AGE levels were identified in RPE cells for isolated samples of AMD patients (69-70). It is known that the increase in AGE and RAGE activates NF- $\kappa\beta$  in RPE cells (70).

Activation of NF- $\kappa\beta$  induces an increase in the expression of several inflammatory cytokines and adhesion molecules that can potentially trigger and/or worsen AMD. Among those to be highlighted are:

# Tumor necrosis factor-α (TNF-α)

TNF- $\alpha$  is a low molecular weight protein, produced, predominantly, by activated macrophages. It has the potential to modulate the production and expression of the vascular endothelial growth factor (VEGF) receptors (71-72). This cytokine may play a cell protective or destructive role. These characteristics may be closely associated with its receptors tumor necrosis factor receptor superfamily member 1A (Tnfrsf1a) and tumor necrosis factor receptor superfamily member 1B (Tnfrsf1b). It is known that activation of Tnfrsf1a induces inflammation, inhibition of the endothelial cellular migration and apoptosis of the endothelial cells (73). Additionally, it has shown potential to inhibit the choroidal neovascularization (CNV). Conversely, Tnfrsf1b receptors regulate lymphocyte proliferation (74) and promote endothelial cell activation, migration, and survival (75-76). In this regard, Tnfrsf1b, unlike Tnfrsf1a, may promote CNV (77). An experimental study reported that TNF- $\alpha$  down-regulates VEGF secretion in polarized RPE cells but up-regulates it in nonpolarized RPE cells. These results are due to the opposing activity levels of the c-Jun N-terminal kinase (JNK) and NF- $\kappa\beta$  pathways. In certain clinical conditions, such as AMD, the RPE cell polarity changes at different stages of the disease with the RPE cells being polarized early on, and some RPE cells losing their cellular polarity at the later stages (36). In the physiopathogenesis of the choroidal neovascular membrane (CNVM), experimental and clinical studies demonstrated that intervention in this cytokine may improve angiogenesis progression (78-80). TNF-α also stimulates the production of Interleukin-6 (81).

#### Interleukin-6 (IL-6)

IL-6 is a multifunctional cytokine that acts upon a wide range of cell tissues and linings. It is considered a potent mediator of the inflammation and immune response (8182) and is a marker for systemic inflammation. Human RPE cells constitutively express and release IL-6 at a relatively low level (83). Several AMD studies have reported IL-6 to be an important regulator of CNV, as it also acts upon VEGF expression (47,84-87). Increase in IL-6 levels were observed in a laser-induced CNV mouse model and the blockage of its receptors induced a significant decrease in the expression of monocyte chemoattractant protein-1 (MCP-1/CCL2), VEGF and inhibited macrophage infiltration into the CNV areas (88). A prospective cohort study demonstrated that elevated IL-6 may serve as marker for the progression of AMD (89). However, another study found no significant association between plasma IL-6 levels and AMD, or AMD progression (90). The inhibition of NF-kB activation decreased the H2O2-induced increase of IL-6 release by RPE cells, demonstrating NF-κB effect on this inflammatory interleukin (91).

#### Inducible Nitric Oxide Synthase (NOS-2 or iNOS)

Nitric oxide synthase (NOS) is a family of enzymes that catalyzes the production of nitric oxide (NO), from Larginine. This family presents three isoforms: NOS-1 or neuronal (nNOS); NOS-2, or inducible or immunological (iNOS); and NOS-3 or endothelial (eNOS). The three isoforms are found in different eye tissues (92,93). Overproduction of the free radical NO has been associated with the pathogenesis of a variety of inflammatory and immunologically mediated diseases as well as with the induction and progression of AMD (94). Activation of NF $k\beta$  induces not only the iNOS expression, but also the pathological conditions caused by the endotoxins or the cytokines such as IL-1, IL-6, and TNF-α. Upon induction, iNOS will produce a large amount of NO for a long period of time (95). In this condition, NO is converted into NO2, nitrite, peroxynitrite and free radicals to induce pathophysiological alterations such as AMD (94-96). It has been demonstrated that a specific NF-kB inhibitor, pyrrolidine dithiocarbonate (PDTC), reduced iNOS expression in RPE cells treated with linoleic acid (LA) (37).

# Interleukin-1β (IL-1β)

IL-1 $\beta$  is a pro-inflammatory cytokine that may initiate innate immunological processes associated with inflammation, infection, and immunity (97-98). In the retina, immunoreactivity to IL-1 has been observed in the astrocytes and Müller cells (99). IL-1 $\beta$  is secreted as an inactive form and requires proteolytic cleavage by the caspase-1 enzyme to be released in an active form (100). Caspase-1 activation platform, known as inflammasome, has been associated with AMD physiopathogenesis (101-102).

A study on AMD experimental models, with geographic atrophy of the choroid, has reported that mononuclear phagocytes express IL-1 $\beta$ , responsible for the lesion of cone outer elements and death of rods (103-105). Another study demonstrated that IL-1 $\beta$  induces rod degeneration through the disruption of retinal glutamate homeostasis (106). Additionally, patients with polypoidal choroidal vasculopathy and wet AMD presented a significant increase in the expression of IL-1 $\beta$  in vitreous (107). Suppression of IL-1 $\beta$  expression by salicin has shown to inhibit activation of NF-k $\beta$  in retinal endothelial cells, representing a potential therapeutic approach to treat CNV in AMD (108).

# Interleukin-2 (IL-2)

IL-2 plays crucial roles in regulating both immune activation and homeostasis (109). It is mainly produced by activated T cells, especially CD4+, and is synthesized in smaller amount by B cells and monocytes. IL-2 is one of the main T-cell stimulating factor (110) and has already been associated with AMD (111). A study has reported an increased activation of the inflammation pathway IL-2, which is consistent with the conclusions drawn from clustering analysis of several AMD phenotype-specific RPE-choroid modules that inflammation is a prevalent functional category (112). Another study investigated the effects of IL-2 on epithelial-mesenchymal transition (EMT), extracellular matrix (ECM) synthesis and transforming growth factor  $\beta 2$  (TGF- $\beta 2$ ) expression in RPE cells. The results indicated that the signal transducer and activator of transcription 3 (STAT3) and NF-κβ signaling pathways might interact with each other and play important roles in IL-2-induced fibrosis in RPE cells together. These findings can offer new insights about the molecular mechanisms underlying the pathogenesis of AMD (113).

# Interleukin-8 (lL-8)

IL-8 was identified in 1987 as a novel type of neutrophilactivating cytokine. It is released by phagocytes and a wide variety of tissue cells upon exposure to inflammatory stimuli (114). IL-8 also promotes an increase in the expression of adhesion molecules by the endothelial cells and activates the polymorphonuclear neutrophils, increasing the oxidative metabolism (115).

A meta-analysis suggested that IL-8 +781 C/T polymorphism affects predisposition to AMD and wet AMD. Moreover, patients with AMD and wet AMD also present elevated IL-8 levels (116). A case-control study suggested a possible secondary role of IL-8 gene in the development of AMD and regarded IL-8 as a new susceptibility genomic biomarker of AMD (117). The intraocular IL-8 concentrations have been elevated in patients with exudative AMD (118) and correlated with the size of an active CNV (119). Studies have shown that NFkß activity is upregulated in the presence of 25hydroxycholesterol (25-OH), a potent inducer of IL-8 expression and secretion in human adult retinal pigment epithelial (ARPE-19), and that this transcription factor is, at least, partially involved in IL-8 production upon 5-OH treatment (120).

# Cyclooxygenase 2 (COX-2)

COX-2 belongs to an enzyme group formed by COX isoforms COX-1, COX-2 and COX-3, which is involved in inflammatory immune responses required for the conversion of arachidonic acid to prostaglandins (121). It mediates inflammation and is induced by pathological stimuli including cytokines, growth factors, inflammatory mediators, and bacterial lipopolysaccharides (121-122). In humans, COX-2 is detected in the outer plexiform layer and in RPE cells (123,124). COX-2 has been shown to modulate the expression of VEGF ligand and its receptors, an important mediator in the development of ocular neovascularization (125). COX-2 involvement has been associated with CNVMs and subretinal fibrosis of the

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retina (125). It has been shown that COX-2 could stimulate macrophages to produce TGF-B, which consequently synthesizes and deposits collagen fibers, eventually leading to fibrosis (126). An experimental study on wet AMD demonstrated that COX-2-selective inhibitor reduces subretinal fibrosis in vivo and in vitro (127). This experiment confirmed the role of COX-2 in the AMD physiopathogenesis. Corroborating the experimental findings, the immunohistochemical analysis of CNVs in humans revealed an expression of COX-2 in 69% of the cases, confirming the theory that inflammation is an important component in the development and progression of neovascular AMD in some patients (128). Research has shown that a specific NF-kB inhibitor, the pyrrolidine dithiocarbonate (PDTC), significantly reduced the expression of COX-2 in RPE cells treated with the LA, a fatty acid involved in AMD genesis, indicating that activation of NF-kB was involved in LA-induced expression of COX-2 (37).

# C-reactive protein (CRP)

CRP is a highly conserved acute phase protein of the pentraxin family that consists of 5 noncovalently linked subunits of  $\approx$ 23 kDa. It is predominantly produced in the liver, although, under certain conditions, it can also be secreted by smooth muscle cells and endothelial cells (129,130).

CRP is released into circulation upon stimulation by IL-6 and other cytokines (131). Several studies suggest a close association between serum CRP and AMD (132-134).

A meta-analysis study showed that high serum levels (> 3 mg/L) of CRP are associated with a two-fold likelihood of late onset AMD, compared to low levels (< 1 mg/L) (135). The Rotterdam study found that elevated baseline levels of highly sensitive CRP were associated with the development of early and late AMD in the large population-based cohort (136).

A study on genotyped human donor eyes reported that eyes homozygous for the high-risk CFH (Y402H) allele had elevated monomeric CRP (mCRP) within the choriocapillaris and Bruch's membrane, compared to those with the low-risk genotype. This study indicated that mCRP is the most abundant form of CRP in human choroid, and that mCRP levels are elevated in individuals with the high-risk CFH genotype. Moreover, proinflammatory mCRP significantly affected endothelial cell phenotypes in vitro and ex vivo, suggesting a substantial role for mCRP in choroidal vascular dysfunction in AMD (137). Significant CRP deposition has shown to trigger and exacerbate the inflammatory response in RPE cells promoted by the induction of pro- inflammatory cytokines such as IL-8. This induction is mediated by NF-kB and multiple Mitogen-activated protein kinase (MAPK) pathways through Fc gamma receptors. Thus, it might contribute to the accumulation of immune cells observed in areas of drusen formation and choroidal neovascularization (138).

#### **E-Selectin**

E-selectin, known as the endothelial leukocyte adhesion molecule 1 (ELAM-1), is responsible for the regulation of the first processes in the adhesion cascade, binding and rolling of leukocytes into the endothelium. E-selectin is inducibly expressed in endothelial cells (139). An immunohistochemistry study demonstrated that subfoveal CNVMs surgically excised from AMD patients presented higher ICAM-1 and E-selectin immunostaining when compared with those in the normal eye and that the increase in ICAM-1 and E-selectin immunoreactivity occurs primarily in the periphery of the CNVMs, where there are larger numbers of vessels. However, this immunoreactivity was not identified on any larger patent vessels in the central, fibrotic regions of the CNVMs (140). An experimental wet AMD study reported an increase in E-selectin in RPE, in the choroidal vascular endothelial and inflammatory cells (141). However, another study was not able to demonstrate an association between the E-selectin single nucleotide polymorphism (SNPs) and AMD development (142).

#### Intercellular Adhesion Molecule 1 (ICAM-1)

ICAM-1 or CD-54 is a glycoprotein of the immunoglobulin superfamily. Like other adhesion molecules, ICAM-1 is distributed in the endothelial cells and leukocytes, participating in the leukocyte recruitment to damaged or inflamed tissue (143). It is known that, in the normal eye, ICAM-1 is expressed in low levels in the choroid and retina vascular endothelium, as well as in the RPE, Bruch's membrane and outer limiting membrane (144-146). It was also demonstrated that ICAM-1 presents a higher concentration in the macular region than in the peripheral region (147). This finding suggests a higher susceptibility of the macula for the traffic of immune cells, including the macrophages, which accounts for the higher incidence of CNV in this region. It is known that the macrophages, besides producing VEGF (26), are also sources of inflammatory and proangiogenic cytokines, which mediate the inflammatory response and contribute, significantly, to the formation of CNVM (89,148-151). In a hypercholesterolaemic experimental model, an increase in the ICAM-1 and interleukin-6 expression in the sclerochoroidal complex was observed (152). In pathological conditions, as well as in AMD, a significant increase in the expression of ICAM-1 in RPE vessels and cells was observed. This increase in immunoreactivity was primarily observed in CNVM periphery, where there are a large number of vessels (153). Other experiments have also reported an increase in the ICAM-1 expression in the RPE, choroidal vascular endothelial and inflammatory cells in wet AMD (144, 154).

A study on CD18-and ICAM-1-deficient mice reported that they developed less CNVM when compared with normal mice, suggesting that this immunoglobulin plays an important role in the formation of CNVM (155). The analysis of the aqueous humor of patients who underwent cataract surgery revealed that concentrations of MCP-1, soluble intercellular cell adhesion molecule-1 (sICAM-1), and soluble intercellular cell adhesion molecule-1 (sVCAM-1) were significantly associated with exudative AMD, even in the presence of normal VEGF concentrations. This study concluded that MCP-1, sICAM-1, and sVCAM-1 could potentially be additional target molecules in the treatment of exudative AMD (156). It has been demonstrated that the expression of ICAM-1 and MMP-9 in ARPE-19 cells may be reduced by quercetin, a flavonoid polyphenolic, via the MEK1/2-ERK1/2 and PKCδ-JNK1/2-c-Jun or NF-κB pathways (157). Another study corroborates this finding by demonstrating that an NF- $\kappa$ B inhibitor (Bay 11-7082)

reduced the expression of ICAM-1, sICAM-1, IL-6, IL-8 and MCP-1 in ARPE-19 cells (158).

#### VCAM-1 (Vascular cell adhesion protein 1)

VCAM-1 is expressed in endothelial cells in response to cytokines (e.g.,  $TNF\alpha$  and  $IL-1\beta$ ) and mediates adhesion of leukocytes including lymphocytes and monocytes (159-160). VCAM-1 is over-expressed in a number of human ocular diseases (161-162).

An experimental study aiming at assessing the role of inflammation as a mechanism of vision loss and degeneration of the sensory retina underlying CNV, reported extensive macrophage recruitment in the retina under CNV. Macrophages were closely associated with retinal blood vessels strongly immunoreactive for VCAM 1, ICAM 1, or platelet-endothelial cell adhesion molecule (PECAM). The macrophage infiltration was responsible for the Müller cell activation, suggesting that macrophages induce degenerative changes in the retina under CNV (163).

A longitudinal population-based cohort study examined the relationship between serum markers of inflammation, oxidative stress, and endothelial dysfunction with a 20year cumulative incidence of early AMD. It reported a modest relationship of serum high-sensitivity CRP, TNF- $\alpha$  receptor 2, and IL-6 to soluble VCAM-1 of early AMD, regardless of age, smoking status, and other factors (164).

#### CONCLUSION

Activation of NF- $\kappa\beta$  induces an increase in the expression of genes associated with inflammatory cytokines, enzymes, and adhesion molecules, which, in turn, are

#### REFERENCES

- Sobrin L, Seddon JM. Nature and nurture- genes and environment- predict onset and progression of macular degeneration. Prog Retin Eye Res. 2014;40:1-15.
- [2] Friedman DS, O'Colmain BJ, Munoz B, Tomany SC, McCarty C, de Jong PT, et al. Prevalence of age-related macular degeneration in the United States. Arch Ophthalmol. 2004;122:564-572.
- [3] Beatty S, Koh H, Phil M, Henson D, Boulton M. The role of oxidative stress in the pathogenesis of age-related macular degeneration. Surv Ophthalmol. 2000;45(2):115-134.
- [4] Masuda T, Shimazawa M, Hara H. Retinal diseases associated with oxidative stress and the effects of a free radical scavenger (Edaravone). Oxid Med Cell Longev. 2017; 9208489.
- [5] Zhu X, Wang K, Zhang K, Zhou F, Zhu L. Induction of oxidative and nitrosative stresses in human retinal pigment epithelial cells by all-trans-retinal. Exp Cell Res. 2016; 348:87-94.
- [6] Coleman HR, Chan CC, Ferris FL, Chew EY. Age-related macular degeneration. Lancet. 2008;372:1835-1845.
- [7] Winkler BS, Boulton ME, Gottsch JD, Sternberg P. Oxidative damage and age-related macular degeneration. Mol Vis.1999;5:32.
- [8] Pauleikhoff D, Harper CA, Marshall J, Bird AC. Aging changes in Bruch's membrane: a histochemical and morphologic study. Ophthalmology.1990; 97:171-178.
- [9] Ruberti JW, Curcio CA, Millican CL, Menco BP, Huang JD, Johnson M. Quick-freeze/deep-etch visualization of age-related lipid accumulation in Bruch's membrane. Invest Ophthalmol Vis Sci. 2003;44:1753-1759.
- [10] Hageman GS, Mullins RF, Russell SR, Johnson LV, Anderson DH. Vitronectin is a constituent of ocular drusen and the vitronectin gene is expressed in human retinal pigmented epithelial cells. FASEB J. 1999;13:477-484.

closely related to AMD. The molecules derived from the activation of this nuclear transcription factor, such as TNF- $\alpha$ , IL-6, IL-8, COX-2, and ICAM-1, have been considered therapeutical targets of experiments related to macular degenerative disease. The other molecules, despite having their role in AMD physiopathogenesis determined, have not, surprisingly, received the same attention. Currently, AMD treatment is predominantly provided with anti-VEGF substances. By analyzing the large number of molecules involved in AMD genesis, mainly those derived from NF- $\kappa\beta$  activation, we may expect that more preventive and therapeutic treatments will be offered in the next decades.

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#### **COMPETING INTERESTS**

The authors declare no competing interests with this case.

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- [11] Mullins RF, Russell SR, Anderson DH, Hageman GS. Drusen associated with aging and age-related macular degeneration contain proteins common to extracellular deposits associated with atherosclerosis, elastosis, amyloidosis, and dense deposit disease. FASEB J. 2000;14: 835-846.
- [12] Hageman GS, Luthert PJ, Victor-Chong NH, Johnson LV, Anderson DH, Mullins RF. An integrated hypothesis that considers drusen as biomarkers of immune-mediated processes at the RPE-Bruch's membrane interface in aging and age-related macular degeneration Prog Retin Eye Res. 2001; 20:705-732.
- [13] Nozaki M, Raisler BJ, Sakurai E, Sarma JV, Barnum SR, Lambris JD, et al. Drusen complement components C3a and C5a promote choroidal neovascularization. Proc Natl Acad Sci. 2006;103:2328-2333.
- [14] Nagineni CN, Samuel W, Nagineni S, Pardhasaradhi K, Wiggert B, Detrick B, et al. Transforming growth factor-β induces expression of vascular endothelial growth factor in human retinal pigment epithelial cells: Involvement of mitogen-activated protein kinases. J Cell Physiol. 2003;197:453-462.
- [15] Penfold PL, Liew S, Madigan MC, Provis JM. Modulation of major histocompatibility complex class II expression in retinas with age-related macular degeneration. Invest Ophthalmol Visual Sci.1997;38(10):2125-2133.
- [16] Penfold PL, Madigan MC, Gillies MC, Provis JM. Immunological and aetiological aspects of macular degeneration. Prog. Retinal Eye Res.2001;20:385-414.
- [17] Killingsworth MC. Age-related components of Bruch's membrane in the human eye. Graefes Arch Clin Exp Ophthalmol. 1987;225:406-412.
- [18] Holtkamp GM, Kijlstra A, Peek R, de Vos AF. Retinal pigment epithelium-immune system interactions: cytokine

production and cytokine-induced changes. Prog. Retinal Eye Res. 2001;20:29-48.

- [19] Loeffler KU, Mangini NJ. Immunolocalization of ubiquitin and related enzymes in human retina and retinal pigment epithelium. Graefe's Arch. Clin. Exp. Ophthalmol.1997; 235:248-254.
- [20] Higgins GT, Wang JH, Dockery P, Cleary PE, Redmond HP. Induction of angiogenic cytokine expression in cultured RPE by ingestion of oxidized photoreceptor outer segments. Invest Ophthalmol Vis Sci. 2003;44:1775-1782
- [21] Yang D, Elner SG, Chen X, Field MG, Petty HR, Elner VM. MCP-1-activated monocytes induce apoptosis in human retinal pigment epithelium. Invest Ophthalmol Vis Sci. 2011;52(8):6026-6034
- [22] Lueck K, Wasmuth S, Williams J, Hughes TR, Morgan BP, Lommatzsch A, et al. Sub-lytic C5b-9 induces functional changes in retinal pigment epithelial cells consistent with age-related macular degeneration. Eye (Lond).2011;25(8):1074-1082.
- [23] Karlstetter M, Ebert S, Langmann T. Microglia in the healthy and degenerating retina:Insights from novel mouse models. Immunobiology 2010;215:685-691.
- [24] Welser-Alves JV, Milner R. Microglia are the major source of TNF-α and TGF-β1 in postnatal glial cultures; regulation by cytokines, lipopolysaccharide, and vitronectin. Neurochem Int. 2013;63:47–53.
- [25] Planck SR, Dang TT, Graves D, Tara D, Ansel JC, Rosenbaum JT. Retinal Pigment Epithelial Cells Secrete Interleukin-6 in Response to Interleukin-1. Invest Ophthalmol Vis Sci. 1992;33:78-82.
- [26] Grossniklaus HE, Ling JX, Wallace TM, Dithmar S, Lawson DH, Cohen C, et al. Macrophage and retinal pigment epithelium expression of angiogenic cytokines in choroidal neovascularization. Mol Vis. 2002;8:119-126.
- [27] Tsutsumi C, Sonoda KH, Egashira K, Qiao H, Hisatomi T, Nakao S, et al. The critical role of ocular-infiltration macrophages in the development of choroidal neovascularization. J Leukoc Biol. 2003;74:25-32.
- [28] Tsutsumi-Miyahara C, Sonoda KH, Egashira K, Ishibashi M, Qiao H, Oshima T, et al. The relative contributions of each subset of ocular infiltrated cells in experimental choroidal neovascularization. Br J Ophthalmol. 2004;88:1217-1222.
- [29] Barnes PJ, Karin M. Nuclear factor-kB A pivotal transcription factor in chronic inflammatory diseases. New Engl. J. Med. 1997; 336: 1066-71.
- [30] Sen CK, Packer L. Antioxidant and redox regulation of gene transcription. FASEB J. 1996;10, 709-720.
- [31] Pantano C, Reynaert NL, van d Vliet A, Janssen-Heininger YM. Redox-sensitive kinases of the nuclear factor-kappaB signaling pathway. Antioxid Redox Signal. 2006;8:1791-1806.
- [32] Schreck R, Baeuerle PA. Assessing oxygen radicals as mediators in activation of inducible eukaryotic transcription factor NF-kappa B. Methods Enzymol. 1994; 234,151-163.
- [33] Schreck R, Albermann K, Baeuerle PA. Nuclear factor kappa B: an oxidative stress-responsive transcription factor of eukaryotic cells (a review). Free Rad Res Commun. 1992; 17, 221-237.
- [34] Traenckner EB, Wilk S, Baeuerle PA. A proteasome inhibitor prevents activation of NF-kB and stabilizes a newly phosphorylated form of IkB -alpha that is still bound to NF-kB. EMBO J. 1994;13, 5433-5441.
- [35] Kretz-Remy C, Mehlen P, Mirault ME, Arrigo AP. Inhibition of I kappa B-alpha phosphorylation and degradation and subsequent NF-kappa B activation by glutathione peroxidase overexpression. J. Cell Biol. 1996;133:1083-1093.
- [36] Terasaki H, Kase S, Shirasawa M, Otsuka H, Hisatomi T, Sonoda S, et al. TNF-a Decreases VEGF Secretion in Highly Polarized RPE Cells but Increases It in Non-Polarized RPE Cells Related to Crosstalk between JNK and NF-kB Pathways. 2013. PLoS ONE 8(7):e69994.
- [37] Fang IM, Yang CH, Yang CM, Chen MS. Linoleic acidinduced expression of inducible nitric oxide synthase and cyclooxygenase II via p42/44 mitogen-activated protein

kinase and nuclear factor-kappaB pathway in retinal pigment epithelial cells. Exp Eye Res. 2007;85(5):667-677.

- [38] Lu H, Lu Q, Gaddipati S, Kasetti RB, Wang W, Pasparakis M, et al. IKK2 Inhibition Attenuates Laser-Induced Choroidal Neovascularization. PLoS ONE. 2014; 9(1): e87530.
- [39] Park H, Lee DS, Yim MJ, Choi YH, Park S, Seo SK. 3,3'-Diindolylmethane inhibits VEGF expression through the HIF-1 $\alpha$  and NF- $\kappa$ B pathways in human retinal pigment epithelial cells under chemical hypoxic conditions Int J Mol Med. 2015;36(1):301-308.
- [40] Xiao W. Advances in NF-κB signaling transduction and transcription. Cell Mol Immunol. 2004;1(6):425-435.
- [41] Sen R, Baltimore D. Multiple nuclear factors interact with the immunoglobulin enhancer sequences. Cell 1986;46:705-716.
- [42] Nabel G, Baltimore D. An inducible transcription factor activates expression of human immunodeficiency virus in T cells. Nature. 1987; 326:711-713.
- [43] Baeuerle PA, Baltimore D. NF-kappa B: ten years after. Cell. 1996; 87: 13-20.
- [44] Baldwin AS. The NF-kappa B and I kappa B proteins: new discoveries and insights. Annu Rev Immunol. 1996 14:649-683.
- [45] Baeuerle PA, Henkel T. Function and activation of NF-kB in the immune system. Annu Rev Immunol. 1994; 12:141-179.
- [46] Pahl HL. Activators and target genes of Rel/NF-kappaB transcription factors. Oncogene 1999;18:6853-6866.
- [47] Salminen A, Ojala J, Huuskonen J, Kauppinen A, Suuronen T, Kaarniranta K. Interaction of aging-associated signaling cascades: inhibition of NF- kappaB signaling by longevity factors FoxOs and SIRT1. Cell Mol Life Sci. 2008; 65, 1049–1058
- [48] Aupperle KR, Bennett BL, Boyle DL, Tak PP, Manning AM, Firesteinet GS. NF-kB regulation by IkB kinase in primary fibroblast-like synoviocytes. J. Immunol. 1999; 163:427-433.
- [49] Morgan MJ, Liu ZG. Crosstalk of reactive oxygen species and NF-kappaB signaling. Cell Res. 2011; 21:103-115.
- [50] Chen CC, Rosenbloom CL, Anderson DC, Manning AM. Selective inhibition of E-selectin, vascular cell adhesion molecule-1, and intercellular adhesion molecule-1 expression by inhibitors of I kappa B- alpha phosphorylation. J Immunol. 1995; 155:3538–3545.
- [51] Siebenlist U. NF-kB/IkB proteins. Their role in cell growth, differentiation and development. Biochim Biophys Acta. 1997;1332:7-13.
- [52] Meffert MK, Baltimore D. Physiological functions for brain NF-κB. Trends in Neurosci. 2005; 28:27-43.
- [53] Chen F, Castranova V, Shi X, Demers LM. New insights into the role of nuclear factor-kappaB, a ubiquitous transcription factor in the initiation of diseases. Clin Chem. 1999; 45:7–17.
- [54] Ghosh S, May MJ, Kopp EB. NF-kB and rel proteins: evolutionary conserved mediators of immune responses. Annu Rev Immunol. 1998;16:225-260.
- [55] Malek R, Borowicz KK, Jargielo M, Czuczwar SJ. Role of nuclear factor kB in the central nervous system. Pharmacological Reports. 2007; 59, 25-33
- [56] Li Z, Nabel GJ. A new member of the I kappaB protein family, I kappaB epsilon, inhibits RelA (p65)-mediated NF-kappaB transcription. Mol Cell Biol. 1997;17:6184-6190.
- [57] Spiecker M, Darius H, Liao JK.. A functional role of I kappa B-epsilon in endothelial cell activation. J Immunol. 2000; 164:3316-3322.
- [58] Alcamo E, Hacohen N, Schulte LC, Rennert PD, Hynes RO, Baltimore D. Requirement for the NF- κB family member RelA in the development of secondary lymphoid organs. J Exp Med. 2002; 195,233-244.
- [59] Zandi E, Chen Y, Karin M. Direct phosphorylation of IkappaB by IKKalpha and IKKbeta: discrimination between free and NF-kappaB-bound substrate. Science. 1998; 281:1360-1363.
- [60] Vaughan S, Jat PS. Deciphering the role of nuclear factorkappaB in cellular senescence. Aging (Albany N.Y.) 2011;3:913-919.

- [61] Zandi E, Rothwart DM, Delhase M, Hayakawa M, Karin M. The IkB kanase comples (IKK) contains two kinase subunits, Ikka and IKKb, necessary for IkB phosphorylation and NF-kB activation. Cell 1997;91:243-252
- [62] Kaltschmidt B, Widera D, Kaltschmidt C. Signaling via NF-kB in the nervous system. Biochem Biophys Acta. 2005;1745, 287-299.
- [63] Geleziunas R, Ferrell S, Lin X, Mu Y, Cunningham ETJr, Grant M, et al. Human T-cell leukemia virus type 1 Tax induction of NF-kappaB involves activation of the IkappaB kinase alpha (IKKalpha) and IKKbeta cellular kinases. Mol. Cell. Biol. 1998;18, 5157-5165.
- [64] Cohen P. The TLR and IL-1 signalling network at a glance. Journal of Cell Science. 2014; 127:2383-2390.
- [65] O'Neill LAJ, Kaltschmidt C. NF-kB: a crucial transcription factor for glial and neuronal cell function. TINS 1997; 20:252-258.
- [66] Schottelius AJ, Mayo MW, Sartor RB, Baldwin Jr AS. Interleukin-10 signaling blocks inhibitor of kappaB kinase activity and nuclear factor kappaB DNA binding. J Biol Chem. 1999; 274:31868-31874.
- [67] Devary Y, Rosette C, DiDonato JA, Karin M. NF-kB activation by ultraviolet light not dependent on a nuclear signal. Science. 1993;261: 1442-1445.
- [68] Cherepanoff S, McMenamin P, Gillies MC, Kettle E, Sarks SH. Bruch's membrane and choroidal macrophages in early and advanced age-related macular degeneration. Br J Ophthalmol. 2010;94:918-925.
- [69] Howes KA, Liu Y, Dunaief JL, Milam A, Frederick JM, Marks A, et al. Receptor for advanced glycation end products and age-related macular degeneration. Invest Ophthalmol Vis Sci. 2004;45, 3713-3720.
- [70] Yamada Y, Ishibashi K, Ishibashi K, Bhutto IA, Tian J, Lutty GA, et al. The expression of advanced glycation endproduct receptors in RPE cells associated with basal deposits in human maculas. Exp. Eye Res. 2006; 82:840-848.
- [71] Ryuto M, Ono M, Izumi H, Yoshida S, Weich HA, Kohno K, et al. Induction of vascular endothelial growth factor by tumor necrosis factor alpha in human glioma cells. Possible roles of SP-1. J Biol Chem. 1996;271:28220-28228.
- [72] Patterson C, Perrella MA, Endege WO, Yoshizumi M, Lee ME, Haber E. Downregulation of vascular endothelial growth factor receptors by tumor necrosis factor-alpha in cultured human vascular endothelial cells. J. Clin. Invest. 1996;98:490-496.
- [73] Hsu H, Shu HB, Pan MG, Goeddel DV. TRADD-TRAF2 and TRADD- FADD interactions define two distinct TNF receptor 1 signal transduction pathways. Cell.1996; 84:299-308.
- [74] Wallach D, Arumugam TU, Boldin MP, Cantarella G, Ganesh KA, Goltsev Y, et al. How are the regulators regulated? The search for mechanisms that impose specificity on induction of cell death and NF-B activation by members of the TNF/NGF receptor family. Arthritis Res. 2002; 4:189-196.
- [75] Pan S, An P, Zhang R, He X, Yin G, Min W. Etk/Bmx as a tumor necrosis factor receptor type 2-specific kinase: role in endothelial cell migration and angiogenesis. Moll Cell Biol. 2002; 22:7512-7523.
- [76] Luo D, Luo Y, He Y, Zhang H, Zhang R, Li X, et al. Differential functions of tumor necrosis factor receptor 1 and 2 signaling in ischemia-mediated arteriogenesis and angiogenesis. Am J Pathol. 2006; 169:1886-1898
- [77] Semkova I, Muether PS, Kuebbeler M, Meyer KL, Kociok N, Joussen AM. Recruitment of Blood-Derived Inflammatory Cells Mediated via Tumor Necrosis Factor-Receptor 1b Exacerbates Choroidal Neovascularization. Invest Ophthalmol Vis Sci. 2011; 52:6101-6108.
- [78] Shi X, Semkova I, Müther PS, Dell S, Kociok N, Joussen AM. Inhibition of TNF-alpha reduces laser-induced choroidal neovascularization. Exp Eye Res. 2006; 83:1325-1334.
- [79] Majka S, McGuire PG, Das A. Regulation of matrix metalloproteinase expression by tumor necrosis factor in a murine model of retinal neovascularization. Invest Ophthalmol Vis Sci. 2002; 43:260-266.

- [80] Theodossiadis PG, Liarakos VS, Sfikakis PP, Vergados IA, Theodossiadis GP. Intravitreal administration of the antitumor necrosis factor agent infliximab for neovascular agerelated macular degeneration. Am J Ophthalmol. 2009;147:825-830.
- [81] Elner VM, Scales W, Elner SG, Danforth J, Kunkel SL, Strieter RM. Interleukin-6 (IL-6) gene expression and secretion by cytokine- stimulated human retinal pigment epithelial cells. Exp Eye Res. 1992;54:361-368.
- [82] Kopf M, Baumann H, Freer G, Freudenberg M, Lamers M, Kishimoto T, et al. Impaired immune and acute-phase responses in interleukin-6-defiecient mice. Nature. 1994;368:339-342.
- [83] Nagineni CN, Detrick B, Hooks JJ. Synergistic effects of gamma interferon on inflammatory mediators that induce interleukin-6 gene expression and secretion by human retinal pigment epithelial cells. Clin Diagn Lab Immunol. 1994;1:569–77.
- [84] Van Snick J. Interleukin-6: an overview. Annu Rev Immunol.1990; 8:253-278.
- [85] Koto T, Nagai N, Mochimaru H, Kurihara T, Izumi-Nagai K, Satofuka S, et al. Eicosapentaenoic acid is antiinflammatory in preventing choroidal neovascularization in mice. Invest Ophthalmol Vis Sci. 2007;48:4328-4334.
- [86] Paimela T, Ryhänen T, Mannermaa E, Ojala J, Kalesnykas G, Salminen A, et al. The effect of 17beta-estradiol on IL-6 secretion and NF- kappaB DNA-binding activity in human retinal pigment epithelial cells. Immunol Lett. 2007;110:139-144.
- [87] Cohen T, Nahari D, Cerem LW, Gera N, Levi B. Interleukin-6 induces the expression of vascular endothelial growth factor. J Biol Chem. 1996; 271:736– 741.
- [88] Izumi-Nagai K, Nagai N, Ozawa Y, Mihara M, Ohsugi Y, Kurihara T, et al. Interleukin-6 receptor-mediated activation of signal transducer and activator of transcription-3 (STAT3) promotes choroidal neovascularization. Am J Pathol. 2007;170,2149-2158.
- [89] Seddon JM, George S, Rosner B, Rifai N. Progression of age-related macular degeneration: prospective assessment of C-reactive protein, interleukin 6 and other cardiovascular biomarkers. Arch Ophthalmol. 2005;123: 774-782.
- [90] Klein R, Klein BE, Knudtson MD, Wong TY, Shankar A, Tsai MY. Systemic markers of inflammation, endothelial dysfunction, and age-related maculopathy. Am J Ophthalmol 2005;140: 35-44.
- [91] Wu WC, Hu DN, Gao HX, Chen M, Wang D, Rosen R, McCormick SA. Subtoxic levels hydrogen peroxideinduced production of interleukin-6 by retinal pigment epithelial cells. Mol Vis. 2010 Sep 12;16:1864-73.
- [92] Behar-Cohen FF, Goureau O, D'Hermies F, Courtois Y. Decreased intraocular pressure induced by nitric oxide is correlated to nitrite production in the rabbit eye. Invest Ophthalmol Vis Sci. 1996; 37:1711-1715.
- [93] Park C-S, Pardhasaradhi K, Gianotti C, Villegas E, Krishna G. Human retina expresses both constitutive and inducible isoforms of nitric oxide synthase mRNA. Biochem Biophys Res Commun. 1994; 205:85-91.
- [94] Ando A, Yang A, Nambu H, Campochiaro PA. Blockade of nitric-oxide synthase reduces choroidal neovascularization. Mol Pharmacol. 2002; 62, 539-544
- [95] Szabo C, Thiemermann C. Regulation of the expression of the inducible isoform of nitric oxide synthase. Adv Pharmacol. 1995; 34;113-153.
- [96] Cantó A, Olivar T, Romero FJ, Miranda M. Nitrosative stress in retinal pathologies: Review. Antioxidants. 2019; 8:543.
- [97] Rock KL, Lai JJ, Kono H. Innate and adaptive immune responses to cell death. Immunological reviews. 2011; 243(1): 191–205.
- [98] Xu J, Yin Z, Cao S, Gao W, Liu L, Yin Y, et al. Systematic review and meta-analysis on the association between IL-1B polymorphisms and cancer risk. PLoS One. 2013; 8(5): e63654.
- [99] Scuderi S, D'Amico AG, Federico C, Saccone S, Magro G, Bucolo C, et al. Different retinal expression patterns of ILlalpha, IL-lbeta, and their receptors in a rat model of type

1 STZ-induced diabetes. J Mol Neurosci. 2015;56:431-439.

- [100] Miao EA, Rajan JV, Aderem A. Caspase-1-induced pyroptotic cell death. Immunological reviews. 2011; 243(1): 206–214.
- [101] Doyle SL, Campbell M, Ozaki E, Salomon RG, Mori A, Kenna PF, et al. NLRP3 has a protective role in age-related macular degeneration through the induction of IL-18 by drusen components. Nat Med. 2012;18(5):791–798.
- [102] Anderson OA, Finkelstein A, Shima DT. A2E induces IL-1ss production in retinal pigment epithelial cells via the NLRP3 inflammasome. PLoS One. 2013; 8(6): e67263.
- [103] Chow A, Brown BD, Merad M. Studying the mononuclear phagocyte system in the molecular age. Nat Rev Immunol. 2011;11:788–798.
- [104] Hu SJ, Calippe B, Lavalette S, Roubeix C, Montassar F, Housset M, et al. Upregulation of P2RX7 in Cx3cr1deficient mononuclear phagocytes leads to increased interleukin- 1beta secretion and photoreceptor neurodegeneration. J Neurosci. 2015; 35:6987–6996.
- [105] Eandi CM, Charles Messance H, Augustin S, Dominguez E, Lavalette S, Forster V, et al. Subretinal mononuclear phagocytes induce cone segment loss via IL-1beta. Elife. 2016;5:e16490.
- [106] Charles-Messance H, Blot G, Couturier A, Vignaud L, Touhami S, Beguier F,et al. IL-1β induces rod degeneration through the disruption of retinal glutamate homeostasis. J Neuroinflammation. 2020;17: 1.
- [107] Zhao M, Bai Y, Xie W, Shi X, Li F, Yang F, et al. Interleukin-1β Level Is Increased in Vitreous of Patients with Neovascular Age-Related Macular Degeneration (nAMD) and Polypoidal Choroidal Vasculopathy (PCV). PLoS ONE. 2015; 10(5): e0125150.
- [108] Song Y, Tian X, Wang X, Feng H. Vascular protection of salicin on IL-1β-induced endothelial inflammatory response and damages in retinal endothelial cells. Artif Cells Nanomed Biotechnol. 2019 Dec;47(1):1995-2002.
- [109] Bayer AL, Pugliese A, Malek TR. The IL-2/IL-2R system: from basic science to therapeutic applications to enhance immune regulation. Immunol Res. 2013 Dec;57(1-3):197-209.
- [110] Hatakeyama M, Tsudi M, Minamoto S, Kono T, Doi T, Miyata T, et al. Interleukin 2 receptor beta chain gene: generation of the three receptor forms by cloned human alpha and beta chain DNAs. Science. 1989;244:551-556
- [111] Makarev E, Cantor C, Zhavoronkov A, Buzdin A, Aliper A, Csoka AB. Pathway activation profiling reveals new insights into age-related macular degeneration and provides avenues for therapeutic interventions. Aging. 2014; 6:1064–1075.
- [112] Newman AM, Gallo NB, Hancox LS, Miller NJ, Radeke CM, Maloney MA, et al. Systems-level analysis of agerelated macular degeneration reveals global biomarkers and phenotype-specific functional networks. Genome Med. 2012; 4:16.
- [113] Jing R, Qi T, Wen C, Yue J, Wang G, Pei C, Ma B Interleukin-2 induces extracellular matrix synthesis and TGF- $\beta$ 2 expression in retinal pigment epithelial cells .Develop Growth Differ. 2019;61:410–418.
- [114] Baggiolini M, Walz A, Kunkcl SL. Neutrophil-activating peptide-1/interleukin 8, a novel cytokine that activates neutrophils. J Clin Invest. 1989; 84,1045-1049.
- [115] Zwahlen R, Walz A, Rot A. In vitro and in vivo activity and pathophysiology of human interleukin-8 and related peptides. Int Rev Exp Pathol. 1993;34:27-42.
- [116] Liu J, Tian Z, Li J, Zhao G. Associations of IL-8 gene polymorphisms and IL-8 levels with predisposition to agerelated macular degeneration: a meta-analysis. Aging Clin Exp Res. 2020; Mar 10.
- [117] Ricci F, Staurenghi G, Lepre T, Missiroli F, Zampatti S, Cascella R, et al. Haplotypes in IL-8 Gene Are Associated to Age-Related Macular Degeneration: A Case- Control Study. PLoS ONE. 2013; 8(6): e66978.
- [118] Jonas JB, Tao Y, Neumaier M & Findeisen P. Cytokine concentration in aqueous humour of eyes with exudative age-related macular degeneration. Acta Ophthalmol. 2012; 90: e381–e388.

- [119] Miao H, Tao Y, Li XX. Inflammatory cytokines in aqueous humor of patients with choroidal neovascularization. Mol Vis. 2012;18: 574–580.
- [120] Catarino S, Bento CF, Brito A, Murteira E, Fernandes AF, Pereira P. Regulation of the expression of interleukin-8 induced by 25-hydroxycholesterol in retinal pigment epithelium cells. Acta Ophthalmol. 2012 Jun;90(4):e255-63.
- [121] Rouzer CA, Marnett LJ. Cyclooxygenases: Structural and functional insights. J Lipid Res. 2009;50:S29–S34.
- [122] Cianchi F, Cortesini C, Bechi P, Fantappiè O, Messerini L, Vannacci Ai, et al. Upregulation of cyclooxygenase 2 gene expression correlates with tumor angiogenesis in human colorectal cancer, Gastroenterology. 2001; 121: 1339– 1347
- [123] Ju WK, Neufeld AH. Cellular localization of cyclooxygenase-1 and cyclooxygenase-2 in the normal mouse, rat, and human retina. J Comp Neurol. 2002;452:392–399.
- [124] Chin MS, Nagineni CN, Hooper LC, Detrick B, Hooks JJ. Cyclooxygenase-2 gene expression and regulation in human retinal pigment epithelial cells. Invest Ophthalmol Vis Sci 2001;42:2338–2346.
- [125] Skold M, Cullheim S, Hammarberg H, Piehl F, Suneson A, Lake S. et al. Induction of VEGF and VEGF receptors in the spinal cord after mechanical spinal injury and prostaglandin administration. Eur J Neurosci 2000;12:3675–3686.
- [126] Connor TB, Roberts AB, Sporn MB, Danielpour D, Dart LL, Michels RG, et al. Correlation of fibrosis and transforming growth factor-beta type 2 levels in the eye. J Clin Investig. 1989;83:1661–1666.
- [127] Zhang R, Liu Z, Zhang H, Zhang Y, Lin D.The COX-2-Selective Antagonist (NS-398) Inhibits Choroidal Neovascularization and Subretinal Fibrosis. PLoS ONE. 2016; 11(1): e0146808.
- [128] Maloney SC, Fernandes BF, Castiglione E, Antecka E, Martins C, Marshall JC, et al. Expression of cyclooxygenase-2 in choroidal neovascular membranes from age-related macular degeneration patients. Retina. 2009; 29: 176–180.
- [129] Calabro P, Willerson JT, Yeh ET. Inflammatory cytokines stimulated C-reactive protein production by human coronary artery smooth muscle cells. Circulation. 2003; 108: 1930–1932.
- [130] Venugopal SK, Devaraj S, Jialal I. Macrophage conditioned medium induces the expression of C-reactive protein in human aortic endothelial cells: potential for paracrine/autocrine effects. Am J Pathol. 2005; 166: 1265– 1271.
- [131] Ganter U, Arcone R, Toniatti C, Morrone G, Ciliberto G. Dual control of C-reactive protein gene expression by interleukin-1 and interleukin-6. EMBO J. 1989 Dec 1; 8(12):3773-3779.
- [132] Čolak E, Kosanović-Jaković N, Žorić L, Radosavljević A, Stanković S, Majkić-Singh N. The association of lipoprotein parameters and C-reactive protein in patients with age-related macular degeneration. Ophthalmic Res. 2011; 46: 125–132.
- [133] Čolak E, Majkić-Singh N, Žorić L, Radosavljević A, Kosanović-Jaković N. The impact of inflammation to the antioxidant defense parameters in AMD patients. Aging Clin Exp Res. 2012; 24: 588–594
- [134] Seddon JM, George S, Rosner B, Rifai N. Progression of age-related macular degeneration: prospective assessment of C-reactive protein, interleukin 6, and other cardiovascular biomarkers. Arch Ophthalmol. 2005 Jun; 123(6):774-782.
- [135] Hong T, Tan AG, Mithchell P, Wang JJ. A review and meta-analysis of the association between C-reactive protein and age-related macular degeneration. Surv Ophthalmol. 2011;56:184-194.
- [136] Boekhoom SS, Vingerling JR, Witteman JC, Hofman A, de Jong P. C-reactive protein level and risk of aging macula disorder: The Rotterdam Study. Arch Ophthalmol 2007;125:1396–1401.
- [137] Chirco KR, Whitmore SS, Wang K, Potempa LA, Halder JA, Stone EM, et al. Monomeric C-reactive protein and

inflammation in age-related macular degeneration. J Pathol. 2016; 240(2): 173–183.

- [138] Wang Y, Bian ZM, Yu WZ, Yan Z, Chen WC, Li XX. Induction of interleukin-8 gene expression and protein secretion by C-reactive protein in ARPE-19 cells. Exp Eye Res. 2010 Aug;91(2):135-42.
- [139] Kannagi R, Izawa M, Koike T, Miyazaki K, I kimura N. Carbohydrate- mediated cell adhesion in cancer metastasis and angiogenesis. Cancer Sci. 2004; 95:377-384.
- [140] Yeh D, Bula D, Miller J, Gragoudas E, Arroyo J: Expression of leukocyte adhesion molecules in human subfoveal choroidal neovascular membranes treated with and without photodynamic therapy. Invest Ophthalmol Vis Sci. 2004, 45:2368-2373.
- [141] Shen WY; Yu MJT; Barry CJ; Constable IJ; Rakoczy PE, Expression of cell adhesion molecules and vascular endothelial growth factor in experimental choroidal neovascularisation in the rat. Brit J Ophthalmol 1998, 82 (9), 1063–1071.
- [142] Bojanowski CM, Tuo J, Chew EY, Csaky KG, Chan C. Analysis of Hemicentin-1, hOgg1, and E-selectin single nucleotide polymorphisms in age-related macular degeneration. Trans Am Ophthalmol Soc. 2005; 103: 37– 45.
- [143] van de Stolpe A, van der Saag PT. Intercellular adhesion molecule-1. J Mol Med. 1996; 74:13-33.
- [144] Duguid IG, Boyd AW, Mandel TE. Adhesion molecules are expressed in the human retina and choroid. Curr Eye Res. 1992;11:153–159.
- [145] Elner SG, Elner VM, Pavilack MA, Todd RF, Mayo-Bond L, Franklin WA et al. Modulation and function of intracellular adhesion molecule-1 (CD54) on human retinal epithelial cells. Lab Invest. 1992;66:200–211.
- [146] McLeod S, Lefer DJ, Merges C, Lutty GA. Enhanced Expression of Intracellular Adhesion Molecule-1 and P-Selectin in the Diabetic Human Retina and Choroid. Am J Pathol 1995; 147:642-653.
- [147] Mullins RF, Skeie JM, Malone EA, Kuehn MH. Macular and peripheral distribution of ICAM-1 in the human choriocapillaris and retina. Mol Vis. 2006;12:224-235.
- [148] May LT, Ghrayeb J, Santhanam U, Tatter SB, Sthoeger Z, Helfgott DC, et al. Synthesis and secretion of multiple forms of beta 2-Interferon/Bcell differentiation factor 2/hepatocyte-stimulating factor by human fibroblasts and monocytes. J Biol Chem.1988; 263:7760-2766.
- [149] Oh H, Takagi H, Takagi C, Suzuma K, Otani A, Ishida K, et al. The potential angiogenic role of macrophages in the formation of choroidal neovascular membranes. Invest Ophthalmol Vis Sci. 1999; 40, 1891–1898.
- [150] Markomichelakis NN, Theodossiadis PG, Sfikakis PP. Regression of neovascular age-related macular degeneration following infliximab therapy. Am J Ophthalmol. 2005;139:537–540.
- [151] Shi X, Semkova I, Muther P S, Dell S, Kociok N, Joussen AM. Inhibition of TNF-alpha reduces laser-induced choroidal neovascularization. Exp. Eye Res. 2006;83:1325–1334
- [152] Torres RJA, Noronha L, Torres RRA, Nagashima S, Torres CLA, Luchini A et al . Increased intercellular adhesion molecule-1 immunoreactivity in the sclera-choroid

complex in hypercholesterolemia experimental model. Rev Bas.Oftalmol. 2014; 73(4): 210-215.

- [153] Deborah C. Yeh, Deisy V. Bula, Joan W. Miller, Evangelos S. Gragoudas, and Jorge G. Arroyo. Expression of Leukocyte Adhesion Molecules in Human Subfoveal Choroidal Neovascular Membranes Treated with and without Photodynamic Therapy. Invest Ophthalmol Vis Sci. 2004;45:2368–2373.
- [154] Nagai N, Oike Y, Izumi-Nagai K, Urano T, Kubota Y, Noda K, et al. Angiotensin II type 1 receptor-mediated inflammation is required for choroidal neovascularization. Arterioscler Thromb Vasc Biol. 2006; 26, 2252–2259
- [155] Sakurai E, Taguchi H, Anand A, Ambati BK, Gragoudas ES, Miller JW, et al. Targeted disruption of the CD18 or ICAM-1 gene inhibits choroidal neovascularization. Invest Ophthalmol Vis Sci. 2003; 44: 2743–2749
- [156] Jonas JB, Tao Y, Neumaier M. Monocyte Chemoattractant Protein 1, Intercellular Adhesion Molecule 1, and Vascular Cell Adhesion Molecule 1 in Exudative Age-Related Macular Degeneration. Arch Ophthalmol. 2010;128(10):1281-1286
- [157] Cheng SC, Wu YH, Huang WC, Pang JS, Huang TH, Cheng CY. Anti-inflammatory property of quercetin through downregulation of ICAM-1 and MMP-9 in TNFα-activated retinal pigment epithelial cells. Cytokine. 2019 Apr;116:48-60.
- [158] Cheng SC, Huang WC, S Pang JH, Wu YH, Cheng CY. Quercetin Inhibits the Production of IL-1β-Induced Inflammatory Cytokines and Chemokines in ARPE-19 Cells via the MAPK and NF-κB Signaling Pathways. *Int J Mol Sci.* 2019;20(12):2957.
- [159] Sawa Y, Sugimoto Y, Ueki T, Ishikawa H, Sato A, Nagato T, et al. Effects of TNF-alpha on leukocyte adhesion molecule expressions in cultured human lymphatic endothelium. J Histochem Cytochem. 2007;55 (7): 721– 733
- [161] Hernandez C; Burgos R; Canton A; Garcia-Arumi J; Segura RM; Simo R, Vitreous levels of vascular cell adhesion molecule and vascular endothelial growth factor in patients with proliferative diabetic retinopathy: a casecontrol study. Diabetes Care. 2001; 24 (3): 516–521.
- [162] Toker E, Kazokoglu H, Sahin S. Cell adhesion molecules in subretinal fluid: Soluble forms of VCAM-1 (vascular cell adhesion molecule-1) and L-selectin. Int Ophthalmol. 1998; 22 (2): 71–76.
- [163] Caicedo A, Espinosa-Heidmann DG, Pina Y, Hernandez EP, Cousins SW. Blood-derived macrophages infiltrate the retina and activate Muller glial cells under experimental choroidal neovascularization. Exp Eye Res. 2005; 81 (1): 38–47.
- [164] Klein R; Myers CE; Cruickshanks KJ, Gangnon RE, Danforth LG, Sivakumaran TA, et all. Markers of Inflammation, Oxidative Stress, and Endothelial Dysfunction and the 20-year Cumulative Incidence of Early Age-related Macular Degeneration: The Beaver Dam Eye Study. JAMA Ophthalmol. 2014 Apr 1;132(4):446-55